



LEGEND MAX[™]
ELISA Kit



Human CCL17 (TARC)

Cat. No. 441107

ELISA Kit for Accurate Quantitation of Human CCL17
(TARC) from Cell Culture Supernatant, Serum, Plasma
and Other Biological Fluids

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It is highly recommended that this manual be read in its entirety before using this product. Do not use this kit beyond the expiration date.

For Research Purposes Only. Not for use in diagnostic or therapeutic procedures. Purchase does not include or carry the right to resell or transfer this product either as a stand-alone product or as a component of another product. Any use of this product other than the permitted use without the express written authorization of BioLegend is strictly prohibited.

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Introduction:

Human CCL17 (TARC), also known as thymus and activation regulated chemokine, was initially isolated from phytohemagglutinin-stimulated peripheral blood mononuclear cells. CCL17 is constitutively expressed in the thymus and, under activation, in several cell types. CCL17-mediated recruitment of Th₂ cells and CLA⁺ CD4⁺ T cells plays a key role in allergic diseases such as atopic dermatitis, allergic asthma, allergic rhinitis, and allergic contact dermatitis. In addition, CCL17 has been detected in idiopathic pulmonary fibrosis. Also, CCL17 and CCL22 secreted by dendrites cells seems to mediate the recruitment of regulatory T cells to sites of inflammation in patients with chronic hepatitis. CCL17 is upregulated under several conditions: TNF- α , IL-4, and IFN- γ in bronchial epithelial cells; TGF- β 1, TNF- α , and IFN- γ in human keratinocyte cell lines; and histamine and prostaglandin E2P in immature DC.

The BioLegend LEGEND MAX™ Human CCL17 (TARC) ELISA Kit is a Sandwich Enzyme-Linked Immunosorbent Assay (ELISA) with a 96-well strip plate that is pre-coated with a polyclonal goat anti-human CCL17 antibody. The detection antibody is a biotinylated polyclonal goat anti-human CCL17 antibody. This kit is specifically designed for the accurate quantitation of human CCL17 from cell culture supernatant, serum, plasma, and other biological fluids. This kit is analytically validated with ready-to-use reagents.

Materials Provided:

Description	Quantity	Volume (per bottle)	Part #
Anti-human CCL17 (TARC) Pre-coated 96-well Strip Microplate	1 plate		77176
Human CCL17 (TARC) Detection Antibody	1 bottle	12 mL	77177
Human CCL17 (TARC) Standard	1 vial	lyophilized	77288
Avidin-HRP D	1 bottle	12 mL	78237
Assay Buffer A	1 bottle	25 mL	78232
Wash Buffer (20X)	1 bottle	50 mL	78233
Substrate Solution F	1 bottle	12 mL	79132
Stop Solution	1 bottle	12 mL	79133
Plate Sealers	4 sheets		78101

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Materials to be Provided by the End-User:

- Microplate reader able to measure absorbance at 450 nm
- Adjustable pipettes to measure volumes ranging from 1 µL to 1,000 µL
- Deionized water
- Wash bottle or automated microplate washer
- Log-Log graph paper or software for data analysis
- Tubes to prepare standard dilutions
- Timer
- Plate Shaker
- Polypropylene vials

Storage Information:

Store unopened kit components between 2°C and 8°C. Do not use this kit beyond its expiration date.

Opened or Reconstituted Components	
Microplate wells	If not all microplate strips are used, remove the excess strips by pressing up from underneath each strip. Place excess strips back in the foil pouch with the included desiccant pack and reseal. Store between 2°C and 8°C for up to one month.
Standard	The remaining reconstituted standard stock solution can be aliquoted into polypropylene vials and stored at -70°C for up to one month. Avoid repeated freeze-thaw cycles.
Detection Antibody	Store opened reagents between 2°C and 8°C and use within one month.
Avidin-HRP D	
Assay Buffer A	
Wash Buffer (20X)	
Substrate Solution F	
Stop Solution	

Health Hazard Warnings:

1. Reagents that contain preservatives may be harmful if ingested, inhaled or absorbed through the skin. Refer to the MSDS online at BioLegend's website for details (www.biolegend.com/msds).
2. Substrate Solution F is harmful if inhaled or ingested. Avoid skin, eye and clothing contact.
3. To reduce the likelihood of blood-borne transmission of infectious agents, handle all serum, plasma and other biological fluids in accordance with NCCLS regulations.

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4. Stop Solution contains strong acid. *Wear eye, hand, and face protection.*
5. Before disposing of the plate, rinse it with an excess amount of tap water.

Specimen Collection and Handling:

Specimens should be clear and non-hemolyzed. If possible, unknown samples should be run at a number of dilutions to determine the optimal dilution factor that will ensure accurate quantitation.

Cell Culture Supernatant: If necessary, centrifuge all samples to remove debris prior to analysis. It is recommended that samples be stored at $< -70^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Serum: Use a serum separator tube and allow clotting for at least 30 minutes, then centrifuge for 10 minutes at $1,000 \times g$. Remove serum layer and assay immediately or store serum samples at $< -70^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Plasma: Collect blood samples in citrate, heparin or EDTA containing tubes. Centrifuge for 10 minutes at $1,000 \times g$ within 30 minutes of collection. Assay immediately or store plasma samples at $< -70^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Reagent and Sample Preparation:

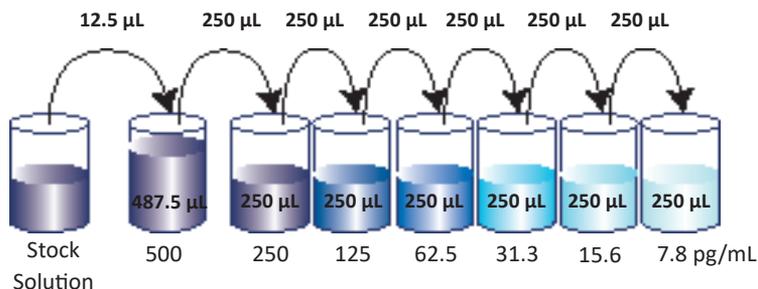
Note: All reagents should be diluted immediately prior to use.

1. Dilute the 20X Wash Buffer to 1X with deionized water. For example, make 1 liter of 1X Wash Buffer by adding 50 mL of 20X Wash Buffer to 950 mL of deionized water. If crystals have formed in the 20X Wash Buffer, bring to room temperature and vortex until dissolved.
2. Reconstitute the lyophilized Human CCL17 (TARC) Standard by adding the volume of Assay Buffer A to make the 20 ng/mL standard stock solution (Refer to LEGEND MAX Kit Lot-Specific Certificate of Analysis/LEGEND MAX Kit Protocol). Allow the reconstituted standard to sit at room temperature for 15-20 minutes, then briefly vortex to mix completely.
3. In general, serum and plasma samples are analyzed without dilutions. However, if dilutions are required, use Assay Buffer A as the sample diluent.
4. Cell culture supernatant samples should be tested initially without dilutions. If dilutions are required, use control culture medium or Assay Buffer A as the sample diluent.

Assay Procedure:

Note: Do not mix reagents from different kits or lots. Reagents and/or antibodies from different manufacturers should not be used with this kit.

1. Bring all reagents to room temperature prior to use. It is strongly recommended that all standards and samples be run in duplicate or triplicate. A standard curve is required for each assay.
2. If not all microplate strips will be used, remove the excess strips by pressing up from underneath each strip. Place excess strips back in the foil pouch with the included desiccant pack and reseal.
3. Prepare 500 μL of the 500 pg/mL top standard by diluting 12.5 μL of the standard stock solution in 487.5 μL of Assay Buffer A. Perform six two-fold serial dilutions of the 500 pg/mL top standard in separate tubes using Assay Buffer A as the diluent. Thus, the human CCL17 standard concentrations in the tubes are 500 pg/mL , 250 pg/mL , 125 pg/mL , 62.5 pg/mL , 31.3 pg/mL , 15.6 pg/mL , and 7.8 pg/mL , respectively. Assay Buffer A serves as the zero standard (0 pg/mL).

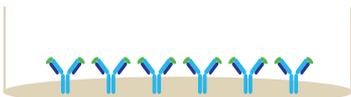
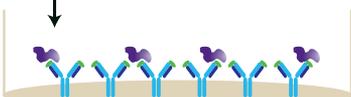
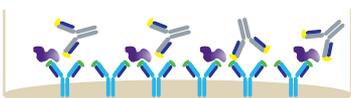
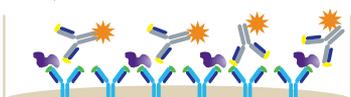
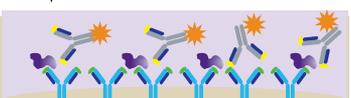


4. Wash the plate 4 times with at least 300 μL of 1X Wash Buffer per well and blot any residual buffer by firmly tapping the plate upside down on absorbent paper. All subsequent washes should be performed similarly.
5. Add 50 μL of Assay Buffer A to each well that will contain either standard dilutions or samples.
6. Add 50 μL of standard dilutions or samples to the appropriate wells
7. Seal the plate with a Plate Sealer included in the kit and incubate the plate at room temperature for 2 hours while shaking at 200 rpm.
8. Discard the contents of the plate into a sink, then wash the plate 4 times with 1X Wash Buffer as in step 4.
9. Add 100 μL of Human CCL17 Detection Antibody solution to each well, seal the plate and incubate at room temperature for 1 hour while shaking.

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10. Discard the contents of the plate into a sink, then wash the plate 4 times with 1X Wash Buffer as in step 4.
11. Add 100 μ L of Avidin-HRP D solution to each well, seal the plate and incubate at room temperature for 20 minutes while shaking.
12. Discard the contents of the plate into a sink, then wash the plate 5 times with 1X Wash Buffer as in step 4. For this final wash, soak wells in 1X Wash Buffer for 30 seconds to 1 minute for each wash. This will help minimize background.
13. Add 100 μ L of Substrate Solution F to each well and incubate for 15 minutes in the dark. Wells containing human CCL17 should turn blue in color with an intensity proportional to its concentration. It is not necessary to seal the plate during this step.
14. Stop the reaction by adding 100 μ L of Stop Solution to each well. The solution color should change from blue to yellow.
15. Read absorbance at 450 nm within 30 minutes. If the reader is capable of reading at 570 nm, the absorbance at 570 nm can be subtracted from the absorbance at 450 nm.

Assay Procedure Summary

1. Wash 4 times
Add 50 μ L Assay Buffer A
A diagram of a well containing immobilized antibodies (blue Y-shapes) on a surface.
2. Add 50 μ L diluted standards or samples
Incubate 2 hr, RT, shaking
A diagram showing antibodies bound to purple Y-shaped standards or samples.
3. Wash 4 times
Add 100 μ L Detection Antibody solution
Incubate 1 hr, RT, shaking
A diagram showing antibodies bound to detection antibodies (grey Y-shapes).
4. Wash 4 times
Add 100 μ L Avidin-HRP D solution
Incubate 20 min, RT, shaking
A diagram showing antibodies bound to Avidin-HRP D solution (orange star-shaped molecules).
5. Wash 5 times
Add 100 μ L Substrate Solution F
Incubate 15 min, RT, in the dark
A diagram showing antibodies bound to substrate solution (purple liquid).
6. Add 100 μ L Stop Solution
A diagram showing antibodies bound to stop solution (yellow liquid).
7. Read absorbance at 450 nm and 570 nm

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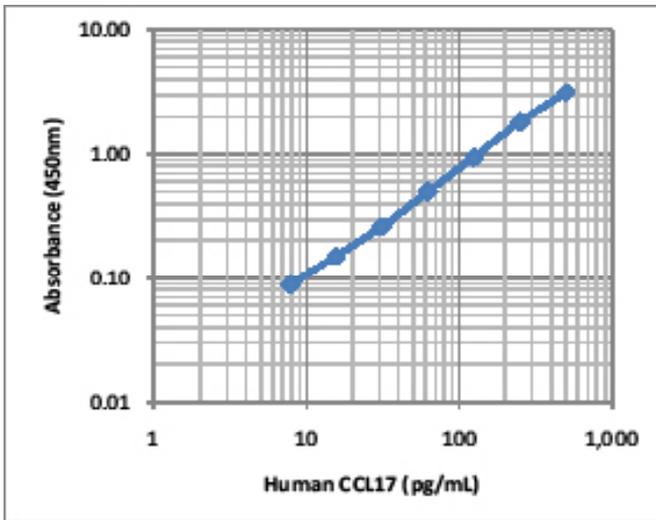
Calculation of Results:

The data can be best calculated with computer-based curve-fitting software using a 5- or 4-parameter logistics curve-fitting algorithm. If an appropriate software is not available, use log-log graph paper to determine sample concentrations. Determine the mean absorbance for each set of duplicate or triplicate standards, controls, and samples. Plot the standard curve on log-log graph paper with chemokine concentration on the X-axis and absorbance on the Y-axis. Draw a best fit line through the standard points. To determine the unknown chemokine concentrations, find the mean absorbance value of the unknown concentration on the Y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the X-axis and read the chemokine concentration.

If samples were diluted, multiply the concentration by the appropriate dilution factor. If a test sample's absorbance value falls outside the linear portion of the standard curve, the test sample needs to be re-analyzed at a higher (or lower) dilution as appropriate.

Typical Data:

This standard curve was generated at BioLegend for demonstration purposes only. A standard curve must be run with each assay.



Performance Characteristics:

Specificity: The antibody pair has negligible cross-reactivity with mouse CCL17, 0.2% when tested at 50 ng/mL. No cross-reactivity was observed when this kit was used to analyze the following recombinant cytokines/chemokines, each at 50 ng/mL.

Human	CCL1, CCL2, CCL3, CCL5, CCL7, CCL8, CCL11, CCL15, CCL19, CCL20, CCL21, CCL22, CCL23, CCL24, CCL25, CCL26, CCL27, CCL28, CXCL1, CXCL2, CXCL3, CXCL5, CXCL6, CXCL7, CXCL9, CXCL10, CXCL11, CXCL12, CXCL13, CXCL17, CX3CL1
Mouse	CCL1, CCL2, CCL7, CCL8, CCL11, CCL20, CCL21, CCL22, CCL24, CCL25, CCL28, CXCL1, CXCL2, CXCL5, CXCL7, CXCL9, CXCL10, CXCL11, CXCL12, CXCL13, CX3CL1, GM-CSF, TACI, TREAK, VEGF

Sensitivity: The minimum detectable concentration of CCL17 is 0.9 ± 0.12 pg/mL.

Recovery: Recombinant human CCL17 was spiked into 8 human serum/plasma samples at concentrations of 250 pg/mL and 62.5 pg/mL, then analyzed with the LEGEND MAX™ Human CCL17 (TARC) ELISA kit. On average, 89.5 % of the chemokine was recovered from the serum samples.

Linearity: Seven human serum/plasma samples spiked with high concentrations of CCL17 were diluted with Assay Buffer A to produce samples with concentrations within the dynamic range and then assayed. On average, 104.6% of the expected chemokine was detected from the serum samples.

Intra-Assay Precision: Sixteen replicates of each of two samples containing different CCL17 concentrations were tested in one assay.

Concentration	Sample 1	Sample 2
Number of Replicates	16	16
Mean Concentration (pg/mL)	255.0	62.0
Standard Deviation	8.0	1.3
%CV	3.1	2.1

Inter-Assay Precision: Two samples containing different concentrations of CCL17 were tested in three independent assays.

Concentration	Sample 1	Sample 2
Number of Assays	3	3
Mean Concentration (pg/mL)	620.3	181.4
Standard Deviation	23.0	9.2
%CV	3.7	5.1

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Biological Samples:

Serum/Plasma - Human serum (n=40) and plasma samples (n = 11) were assayed for natural human CCL17.

Sample Type	Mean (pg/mL)	Range (pg/mL)
Serum (n=40)	60	15-315
EDTA plasma (n=3)	73	41-94
Heparin plasma (n=4)	87	40-126
Citrate plasma (n=4)	62	41-105

Cell Culture Supernates - Human peripheral blood mononuclear cells at a concentration of 2×10^6 cells/mL were co-stimulated with 10 μ g/mL CD3 and 1 μ g/mL CD28 at 37°C for 2 days. The cell culture supernates were collected and assayed for the concentration of natural human CCL17. The concentration of human CCL17 was 6,300 pg/mL in CD3/CD28 stimulated samples, and undetectable in unstimulated samples.

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Troubleshooting Guide:

Problem	Probable Cause	Solution
High Background	Background wells were contaminated	Avoid cross-well contamination by using the provided plate sealers. Use multichannel pipettes and change tips between pipetting samples and reagents.
	Insufficient washes	Increase number of washes. Increase soaking time between washes prior to addition of substrate solution.
	TMB Substrate Solution was contaminated	TMB Substrate Solution should be clear and colorless prior to addition to wells. Use a clean container prior to pipetting substrate solution into wells.
No or poor signal	Detection Antibody, Avidin-HRP or Substrate solution were NOT added	Rerun the assay and follow the protocol.
	Wrong reagent or reagents were added in wrong sequential order	
	Insufficient plate agitation	The plate should be agitated during all incubation steps using a plate shaker at a speed where solutions in wells are within constant motion without splashing.
	The wash buffer contains Sodium Azide (NaN ₃)	Avoid Sodium Azide contamination in the wash buffer as it inhibits HRP activity.
	Incubations were done at an inappropriate temperature, timing or without agitation	Rerun the assay and follow the protocol.
Low or poor standard curve signal	The standard was incorrectly reconstituted or diluted	Adjust the calculations and follow the protocol.
	Standard was inappropriately stored	Store the reconstituted standard stock solution in polypropylene vials at -70°C. Avoid repeated freeze-thaw cycles.
	Reagents added to wells with incorrect concentrations	Check for pipetting errors and the correct reagent volume.

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Problem	Probable Cause	Solution
Signal is high, standard curves have saturated signal	Standard reconstituted with less volume than required	Reconstitute new lyophilized standard with the correct volume of solution recommended in the protocol.
	Standards/samples, detection antibody, Avidin-HRP or substrate solution were incubated for too long	Rerun the assay and follow the protocol.
Sample readings are out of range	Samples contain no or below detectable levels of the analyte	If samples are below detectable levels, it may be possible to use a larger sample volume. Contact technical support for appropriate protocol modifications.
	Samples contain analyte concentrations greater than highest standard point	Samples may require dilution and analysis.
High variation in samples and/or standards	Multichannel pipette errors	Confirm that pipette calibrations are accurate.
	Plate washing was not adequate or uniform	Ensure pipette tips are tightly secured. Ensure uniformity in all wash steps.
	Non-homogenous samples	Thoroughly mix samples before assaying.
	Samples may have high particulate matter	Remove particulate matter by centrifugation.
	Cross-well contamination	Do not reuse plate sealers. Always change tips for reagent additions. Ensure that pipette tips do not touch the reagents on the plate.

ELISA Plate Template

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												



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